








Original Article

Frequency and clinicopathologic features of DNA mismatch repair protein deficiency in colorectal carcinoma in the Turkish population

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Abstract

Objective: Microsatellite instability pathway caused by loss of DNA “mismatch repair genes” (MMR) is responsible for Lynch syndrome-related tumors and 10-15% of sporadic colorectal cancers. Although the MSI test is regarded as the golden standard for the detection of “Lynch syndrome-related tumors”, there is increasing evidence on similar analytic sensitivity of immunohistochemical evaluations.

Methods: We retrospectively evaluated 1,002 colorectal tumors for loss of DNA MMR protein (MLH1, PMS2, MSH2, MSH6) immunohistochemically. The results were correlated with clinicopathological features and high level-microsatellite instability (MSI-H) related histological parameters.

Results: MMR protein expression loss was observed in 9.8% of the cases. MLH1-PMS2 loss (53.2%) was the most common loss, followed by MSH2-MSH6 (31.6%), isolated PMS2 loss (12%), and isolated MSH6 loss (2%). MMR deficiency was more frequent under 50 years old ($p<0.0001$), in right colon tumors ($p<0.0001$), poorly differentiated tumors ($p<0.0001$), tumors with tumor-infiltrating lymphocytes ($p<0.0001$), mucinous component ($p=0.001$), and medullary component ($p<0.0001$). Also, MMR deficiency was less frequent in tumors with tumor budding ($p<0.0001$) and dirty necrosis ($p<0.0001$). The five years-survival rate was 55.7%. No significant correlation was found between MMR deficiency and survival.

Conclusion: MMR deficiency was observed in 9.8% of the cases with distinct clinicopathological features. The results were consistent with previous studies. Unlike the literature, we did not find any statistically significant difference between MMR deficiency and prognosis.

Keywords: Colorectal cancer, microsatellite instability, mismatch repair deficiency, prognosis.



INTRODUCTION

Microsatellite instability (MSI) pathway, which is one of the colorectal carcinogenesis pathways, accounts for 15-20% of sporadic colorectal cancers (CRC) and tumors related to the hereditary nonpolyposis colorectal cancers (HNPCC-Lynch Syndrome) (1,2). Lynch syndrome is an autosomal dominant syndrome characterized by a mutation in DNA mismatch repair (MMR) genes accounting for 2-5% of all colorectal cancers (2,3). There is also an increased incidence of endometrial, renal pelvic, small bowel, ureter tumors and CRC. Since the syndrome is characterized by the development of synchronous and metachronous tumors at an early age, affecting multiple generations of a family, the recognition of an individual with Lynch syndrome enables screening of other family members before the cancer development. It allows more efficient patient management (4). Many guidelines have recommended using PCR-based MSI testing as the gold standard method for selecting patients who undergo further genetic testing concerning Lynch syndrome (2,5). However, MSI testing is not an appropriate method for screening purposes as the test is costly and time-consuming. For this reason, recent studies have compared the analytical sensitivity of the loss of expression in MMR proteins identified by immunohistochemical (IHC) analysis with that of PCR-based MSI testing and reported similar sensitivity rates (6-8). In light of these data, identifying the loss of expression of MMR proteins using IHC methods has taken the first step in the diagnostic algorithm for Lynch syndrome. Another advantage of IHC analysis is its ability to directly identify the expression or loss of expression of the MMR protein, thereby allowing mutation analysis to be focused directly on the gene that leads to protein loss (1). Additionally, some studies propose reflex testing with MMR immunohistochemistry because this finding may play a role in predicting response to therapy and prognosis. Moreover, loss of MMR protein expression is also observed in sporadic cases of colorectal cancer (2,9). The identification of MSI is also crucial as the presence of MSI is a prognostic marker in terms of survival and is of predictive value in the selection of chemotherapy. Many studies to date have found that sporadic MSI tumors are associated with better prognosis and improved overall survival than microsatellite stable (MSS) tumors (10,11). On the other hand, it is reported that high-level microsatellite instability (MSI-H) tumors often poorly respond to 5-fluorouracil (5-FU)-based chemotherapy used in the classical treatment of CRC (5,12). Also, it was established in recent years that individuals with tumors displaying loss of MMR protein expression benefit from the blockage of the programmed cell death protein-1 (PD-1) (13,14). For this reason, the identification of MSI status has an important place in the patient-centered selection of the treatment modality before proceeding with chemotherapy.

The current study assesses histological or clinical parameters (such as age, gender, and localization) that could potentially predict MMR protein deficiency. It also investigates whether the findings of immunohistochemical studies define a patient profile suitable for further MSI testing. In the scope of the present study, clinicopathological data (age, gender, tumor localization, tumor diameter), histological data (tumor type, tumor grade, pT, pN, number of metastatic lymph nodes, LVI, PNI, presence of satellite nodule), and histological data related to the MSI-H phenotype (TIL, Crohn's-like lymphoid reaction, mucinous tumor component, signet ring cell tumor component, medullary tumor component, presence of dirty necrosis, presence of tumor budding, infiltrative tumor margins) were evaluated in 1,002 patients.

MATERIALS AND METHODS

Study group: A total of 1,002 patients who were operated on for colorectal adenocarcinoma and whose resection material was examined were included in the study. Patients with rectal tumors who received neoadjuvant chemotherapy were excluded from the study. The patients' clinicopathological and macroscopic data were retrieved from the hospital's information system, and survival data were obtained from the database of the Cancer Registry Center of the Department of Cancer (CRCDC) in İzmir Public Health Directorate. The histopathological analysis consisted of a retrospective examination of

hematoxylin-eosin (HE)-stained sections of formalin-fixed and paraffin-embedded tissue block.

Histological characteristics: The depth of invasion (pT) and the number of metastatic lymph nodes (pN) were recorded according to the eighth edition of the AJCC-UICC TNM classification (15). Histological findings were re-assessed according to the 2019 recommendations of the World Health Organization (16). Histological tumor type, differentiation, satellite tumor focus, lymphovascular invasion (LVI), and perineural invasion (PNI) were evaluated.

Phenotypic characteristics of MSI-H tumors: The characteristics to be evaluated for MSI-H tumor risk were determined according to the Bethesda criteria and the data collected from the literature (17).

Tumor-infiltrating lymphocytes (TIL): Tumor-infiltrating lymphocytes were evaluated by observing lymphocytes with a halo and a small, blue nucleus in H&E-stained tissue sections. The mean value was calculated by evaluating the number of TIL in ten microscopic high-power fields (HPF). The presence of two or more TIL in an individual HPF was considered to indicate a positive result for MSI-H phenotype (18,19).

Crohn's-like lymphoid reaction: The presence of two or more lymphoid aggregates in tumor-infiltrated borders was considered to indicate a positive reaction (19).

Mucinous differentiation: The tumor was categorized as "mucinous adenocarcinoma" if the extracellular mucin rate exceeded 50%. If the rate was less than 50%, it was classified as a tumor displaying "focal mucinous differentiation" (Figure 1A) (19).

Signet ring cell differentiation: The tumors containing more than 50% signet ring cells were classified as "signet ring cell carcinoma", and the tumors containing less than 50% signet ring cells were classified as "focal signet ring cell differentiation"(Figure 1B) (19).

Poorly differentiated histological subtypes associated with MSI-H: The tumor was considered to exhibit MSI-H phenotype if histological subtypes containing medullary components, and poorly differentiated components exhibiting less than 5% gland formation comprised more than 10% of the tumor (Figure 1C) (16).

Infiltrative tumor margins: Infiltrative tumor margins were examined in a low power field (X4 magnification) and evaluated in two categories: expansive and infiltrative growth patterns.

Tumor budding: The evaluation included assessing the presence of single tumor cells or small cell clusters, each comprising fewer than five cells, exhibiting an anaplastic character at the infiltrative tumor margin, which was independent of the primary tumor mass (Figure 1D) (20).

Dirty necrosis: Dirty necrosis was considered to be positive in the presence of more than 10% cell debris and "dirty" necrosis containing inflammatory cells. The presence of geographic necrosis related to the rapid growth of the tumor was not taken into consideration (19).

Immunohistochemistry: Immunohistochemical analysis involved the examination of a tissue microarray of 5 mm in diameter prepared from H&E-stained tissue sections exhibiting an average tumor area. All IHC staining was performed using a fully automated IHC staining device (Ventana BenchMark XT, Ventana Medical Systems, Tucson, AZ). MLH1 (Clone: ES05 Novacastra; Dilution: 1/150), MSH2 (Clone: 25D12 Novacastra; Dilution: 1/75), MSH6 (Clone: PU29 Novacastra; Dilution:1/50), and PMS2 (Clone: M0R4G Novacastra; Dilution: 1/75) primer antibodies were used in IHC examination. Nuclear staining for each antibody was considered a positive reaction during examination, and the heterogeneity of staining was separately evaluated. A lack of staining in all tumor cells or less than 1% positive nuclear

staining was considered to indicate a loss of "mismatch repair gene" protein.

Evaluation of survival data: Survival analysis involved the assessment of "overall survival" from the date of operational diagnosis to the last control visit on March 2020 for survivors and from the date of diagnosis to death in "months" for nonsurvivors. Early postoperative deaths in the first three months were excluded.

Statistical analysis

All analyses were performed using the SPSS program (version 22.0, SPSS Inc., Chicago, IL, USA). The patients' clinicopathological data and MMR immunohistochemistry results were evaluated using the frequency analysis, and survival was evaluated using Kaplan-Meier survival analysis. Nonparametric tests were used to compare the loss of MMR protein expression with clinicopathological data and survival (Pearson's chi-square and Fisher's exact test, where appropriate), and a Mann-Whitney U test was used to compare age, tumor diameter, number of metastatic LN, and TIL count with other data. A p-value of less than 0.05 was considered statistically significant.

RESULTS

Clinicopathological data: The average age of 1,002 cases included in the study was 63 ± 12.7 years in the range of 22-94 years. Also, at the time of diagnosis, 18% (n=180) of the patients were under 50 years. When the gender distribution of the cases was analyzed, 603 (60.2%) were male, and 399 (39.8%) were female. Furthermore, the mean age at diagnosis was lower in women than in men (61 ± 13.7 versus 65 ± 11.9). Regarding tumor location, 445 (45.4%) of the tumors were located in the rectum, 324 (32.3%) in the left colon, and 223 (22.3%) in the right colon. When rectal tumors were included in left colon tumors, the rate of tumors localized in the left colon was 77.7% (n=779). There were multiple tumors in 64 cases (6.4%), and of these tumors, 36 (56.3%) were diagnosed as synchronous tumors at the time of initial diagnosis, and 28 (43.8%) were diagnosed as metachronous tumors at least six months after surgery. The mean tumor diameter in the resection materials was 5 ± 2.3 cm (range: 1-26 cm). The diameter of the second tumor was not separately evaluated in patients with multiple tumors. It was found that the diameter of the primary tumors localized in the right colon tended to be larger than the diameter of tumors localized in the left colon (6.13 ± 2.7 vs. 5.1 ± 2.2 , $p < 0.05$). The distribution of parameters related to histological features of tumors and phenotypic characteristics features of MSI-H tumors are listed in Table 1 and Table 2, respectively.

Results on loss of MMR protein expression: A loss of expression in one or more MMR proteins was observed in 98 (9.8%) out of 1,002 patients included in the study. Of these patients, 52 (5.2%) had a deficiency in MLH1-PMS2 expression, 31 (3.1%) had a deficiency in MSH2-MSH6 expression, 12 (1.2%) had isolated PMS2 deficiency, and 2 (0.2%) had isolated MSH6 deficiency. Only one patient had a deficiency in MLH1-PMS2 and MSH6 expression.

Heterogeneity: The percentage and intensity of staining were heterogeneous in 40.9% (n=410) of the tumors. The extent of heterogeneity was the highest in the expression of MSH-6 (19%) and PMS2 (15%).

The relationship between the loss of MMR protein expression and clinicopathological, histological, and MSI-H-related findings: Although the rate of loss of MMR protein expression was higher in men, the difference was not statistically significant. The rate of the loss of MMR protein expression was significantly higher in cases under 50 years (18.3% vs. 8.1%; $p < 0.0001$). MMR-deficient tumors were more often localized in the right colon (63.3% vs. 36.7%; $p < 0.0001$) and had a larger diameter (6 ± 3.2 vs. 4.7 ± 2 ; $p < 0.0001$). MMR-deficient tumors were mostly composed of poorly differentiated high-grade

tumors ($p < 0.0001$). The rate of MMR protein deficiency was 31.6% among high-grade tumors and 7.4% among low-grade tumors ($p < 0.0001$). Furthermore, MMR protein deficiency was significantly more common in pT4 and pN0 tumors (16.9%, $p = 0.05$; 8.1%, $p < 0.0001$, respectively). The presence of a satellite nodule was uncommon in MMR-deficient tumors (10.6% vs. 5.8%; $p = 0.05$). The presence of TIL was more common in MMR-deficient tumors ($p < 0.0001$). The rate of Crohn's-like lymphoid reaction was significantly higher in MMR-deficient tumors (13% vs. 7.8%; $p = 0.005$). The rates of focal mucinous differentiation and mucinous adenocarcinoma were significantly higher in cases with MMR-deficient tumors (15.5% and 12.4% vs. 7.3%, $p = 0.001$). Similarly, the rate of medullary tumor component was significantly higher in cases with MMR-deficient tumors (44% vs. 8%, $p < 0.0001$). The rates of tumor budding (4.2% vs. 11.5%, $p = 0.001$) and dirty necrosis (6.4% vs. 13.8, $p < 0.0001$) were significantly lower in MMR-deficient tumors. The relationship between the loss of MMR protein expression and clinicopathological data is presented in Table 3.

Histopathological characteristics of MLH1-PMS2-deficient tumors: MLH1-PMS2-deficient tumors were more commonly localized in the right colon ($p < 0.0001$). The loss of MLH1-PMS2 protein expression was significantly more common in poorly differentiated tumors and tumors with mucinous and medullary components ($p < 0.0001$ for each). The rate of TIL was also higher ($p < 0.0001$). The rates of tumor budding and dirty necrosis were lower ($p = 0.007$ and $p < 0.0001$, respectively) (Figure 2).

Histopathological characteristics of MSH2-MSH6-deficient tumors: MSH2-MSH6-deficient tumors were often localized in the right colon ($p < 0.0001$). The rate of MSH2-MSH6 deficiency was significantly higher in tumors with a medullary component and lower in the presence of tumor budding ($p < 0.0001$ and $p = 0.008$, respectively) (Figure 3). No significant relationship was found with histopathological parameters in tumors with isolated PMS2 and isolated MSH6 deficiency.

Table 1. Histological findings of the cases

	n	%		N	%
Tumor types			pN		
Adenocarcinoma	898	89.6	N0	480	47.9
Mucinous	98	9.8	N1	297	29.6
Signet ring cell carcinoma	6	0.6	N1c	38	3.8
			N2	179	17.9
Tumor differentiation			Lymphovascular invasion		
Well-differentiated	94	9.4	Present	251	25
Moderately differentiated	727	72.6	Absent	751	75
Poorly differentiated	79	7.9			
Tumor grade			Perineural invasion		
Low grade	821	91.2	Present	235	23.5
High grade	79	8.8	Absent	767	76.5
pT			Satellite tumor focus		
T1	15	1.5	Present	173	17.3
T2	99	9.9	Absent	829	82.7
T3	799	79.7			
T4	89	8.9			

Abbreviations: pT: Depth of invasion, pN: Number of metastatic lymph nodes.

	N	%		N	%
TIL			Medullary component		
<2	898	89.6	Present	50	5
>2	104	10.4	Absent	952	95
Crohn's-like reaction			Dirty necrosis		
Present	385	38.4	Present	544	54.3
Absent	617	61.6	Absent	458	45.7
Mucinous component			Tumor budding		
<%50	238	23.8	Present	238	23.8
>%50	105	10.5	Absent	764	76.2
Absent	659	65.8			
Signet ring cell carcinoma component			Tumor border		
Present	39	3.9	Expansive	71	7.1
Absent	963	96.1	Infiltrative	931	92.8

Abbreviations: MSI-H: High level-microsatellite instability, TIL: Tumor-infiltrating lymphocyte.

	MMR deficient CRC	MMR stable CRC	p value
	n=98 (9.8%)	n=904 (90.2%)	
Gender			0.08
Female	46 (46.9%)	353 (39%)	
Male	52 (53.1%)	551 (61%)	
Localization of tumor			<0.0001
Left side	36 (36.4%)	744 (82.3%)	
Right side	62 (63.3%)	160 (17.7%)	
Tumor differentiation			<0.0001
Well	7 (7.1%)	87 (9.6%)	
Moderate	54 (55.1%)	673 (74.4%)	
Poor	25 (25.5%)	54 (6%)	
Mucinous	12 (12.2%)	90 (10%)	
pT stage			0.005
T1	2 (2.0%)	13 (1.4%)	
T2	5 (5.1%)	94 (10.4%)	
T3	76 (77.6%)	723 (80.0%)	

T4	15 (15.3%)	74 (8.2%)	
pN stage			<0.0001
N0	59 (60.2%)	421 (46.6%)	
N1	24 (24.5%)	273 (30.2%)	
N1c	2 (2%)	36 (4%)	
N2	9 (9.2%)	170 (18.8%)	
Nx	4 (4%)	4 (0.4%)	
LVI			0.502
Present	24 (27.8%)	227 (25.1%)	
Absent	74 (75.5%)	677 (74.9%)	
PNI			0.593
Present	17 (17.3%)	218 (24.1%)	
Absent	81 (82.7%)	686 (75.9%)	
Satellite nodule			0.03
Present	10 (10.2%)	163 (18%)	
Absent	88 (89.6%)	741 (82%)	
TIL			<0.0001
Present	25 (25.5%)	79 (8.7%)	
Absent	73 (74.5%)	825 (91.3%)	
Crohn-like reaction			0.005
Present	50 (51%)	335 (37.1%)	
Absent	48 (49%)	569 (62.9%)	
Tumor budding			<0.0001
Present	10 (10.2%)	228 (25.2%)	
Absent	88 (89.8%)	676 (74.8%)	
Dirty necrosis			<0.0001
Present	35 (35.7%)	509 (56.3%)	
Absent	63 (64.3%)	395 (43.7%)	
Tumor border			0.391
Expansive	8 (8.2%)	63 (7%)	
Infiltrative	90 (91.8%)	841 (93%)	

Abbreviations: IHC: Immunohistochemistry, MMR: Mismatch repair, CRC: Colorectal cancer, pT: Depth of invasion, pN: Number of metastatic lymph nodes, LVI: Lymphovascular invasion, PNI: Perineural invasion, TIL: Tumor-infiltrating lymphocytes.

Table 4. Evaluation of MMR deficient CRC and MMR stable CRCs in terms of overall survival

	Estimate time (month) Mean	Std error (month)	95% Confidence interval		p value
			Lower bound	Upper bound	
					0.264
Overall survival (5 years)	101.077	2.488	96.201	105.952	
MMR deficient CRC	102.270	6.936	88.676	115.864	
MMR stable CRC	100.219	2.609	95.201	105.331	

Abbreviations: MMR: Mismatch repair, CRC: Colorectal cancer.

The evaluation was made using the Kaplan-Meier analysis.

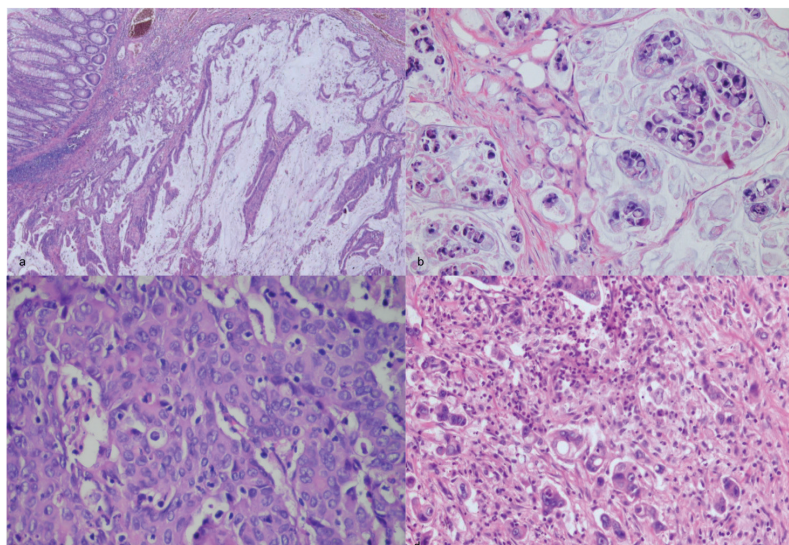


Figure 1. **A)** The morphologic features of the tumor that most determined MMR status in colon adenocarcinoma. MMR deficient mucinous carcinoma with large areas of extracellular mucin and cellular lymphocytic nodules in the surrounding fibrosis, **B)** MMR deficient signet ring cell carcinoma that contained more than 50% signet ring cells, **C)** Tumor-infiltrating lymphocytes in medullary carcinoma that was a rare subtype of poor differentiation tumors, **D)** Tumor budding is the presence of single tumor cells or small cell clusters of less than five cells with an anaplastic character at the infiltrative tumor margin (H&E.)

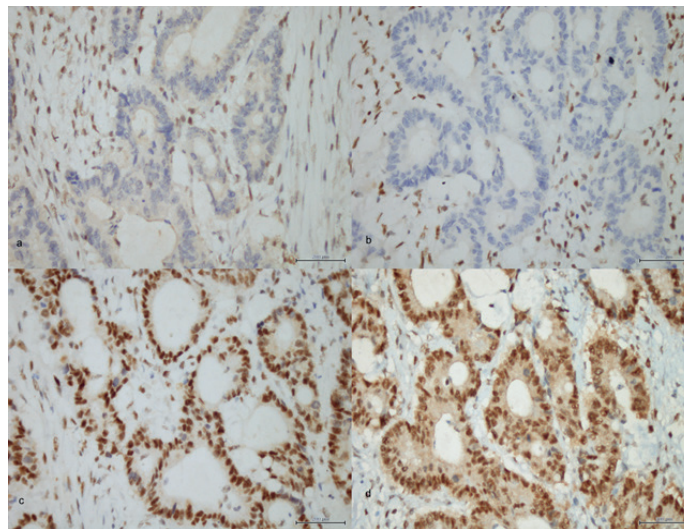


Figure 2. Mismatch repair immunohistochemistry: Loss of MLH1-PMS2 protein expression in moderately differentiated adenocarcinoma cells while stromal lymphocytes show nuclear positivity. Immunohistochemical staining indicates loss of expression MLH1 **A)** with PMS2 **B)** and expression of MSH2 **C)** with MSH6 **D)**.

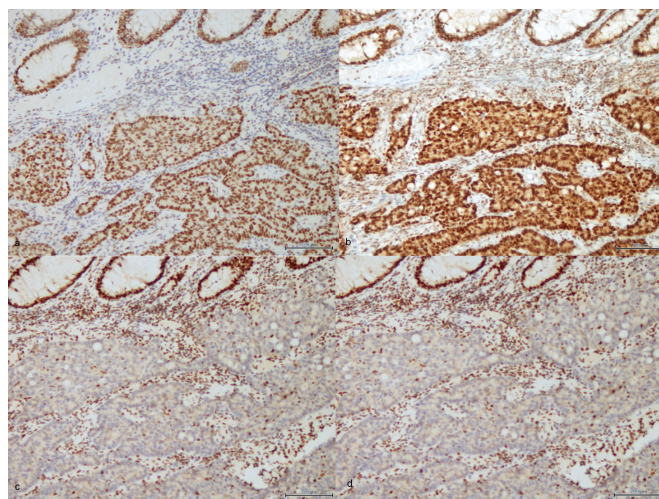


Figure 3. Mismatch repair immunohistochemistry: Loss of MSH2-MSH6 protein expression in poorly differentiated adenocarcinoma cells while normal colon crypts and stromal lymphocytes show nuclear positivity. Immunohistochemical staining indicates expression of MLH1 **A)** with PMS2 **B)** and loss of expression MSH2 **C)** with MSH6 **D)**.

Survival: The analysis of 942 patients, for whom overall survival data were available, showed that 48% of them survived. The overall survival rate was 47.9% among patients with a loss of MMR protein expression versus 43.9% among patients without a loss of MMR protein expression. However, no statistically significant relationship was found between the loss of MMR protein expression and overall survival. The relationship between the loss of MMR protein expression and overall survival is presented in Table 4.

DISCUSSION

IHC revealed a loss of expression in one or more MMR proteins in 98 cases (9.8%). Various studies have reported a rate of MMR protein deficiency as high as 20% in sporadic colorectal cancers (21,22).

Consistent with the literature data, the majority of patients with DNA MMR deficiency were composed of patients with MLH1-PMS2 deficiency (1). The loss of MSH2 expression is often the result of germline mutations, while the loss of MLH1 expression can result from either germline mutations or somatic hypermethylation; thus, MLH1 is one of the most frequent deficiencies (23). One of the patients was evaluated with repeat IHC analysis, which revealed combined MLH1-PMS2 and MSH6 deficiency. This patient was a female older than 50 years old and had a moderately differentiated tumor with an expansive growth pattern. The patient exhibited only Crohn's-like lymphoid reaction, among other phenotypic characteristics of MSI-H. According to a recent theory, a secondary mutation occurs in some individuals with Lynch syndrome, resulting in an extraordinary immunohistochemical staining pattern (24). For example, a somatic mutation can occur in the MSH6 gene over time in a carrier of hereditary mutation in the MLH1-PMS2 gene. These abnormal combinations are generally identified in MSH2-MSH6-PMS2 proteins on rare occasions, and it was asserted that these combinations are independent of the carcinogenesis process (7). It was considered that a similar pathway is involved in this particular case, while there is a need for further molecular evidence. The age at tumor diagnosis was significantly lower in patients with a loss in MMR protein expression than in patients without MMR deficiency. This finding is in agreement with the previous studies reporting a younger age in patients with tumors related to Lynch syndrome than in patients with sporadic colorectal tumors (21,25). The age is often in the range of 50 to 74 years in patients with sporadic tumors exhibiting a loss of MMR protein expression (3,10,21). In our series, among patients younger than 50 years, MMR protein deficiency was detected in 18.3% of cases, which was significantly higher than the rate observed in older patients with MMR protein deficiency. Numerous studies have reported larger diameters in MMR-deficient tumors (11,28). The present study also found a larger tumor diameter in association with the loss of MMR protein expression. It seems reasonable that the finding of MSI tumors with MMR protein deficiency being more often localized in the right colon can be related to larger tumor diameter (9,21,25). The localization of the tumor in the right colon was remarkable in MMR-deficient tumors. The rate of right colon tumors is lower than that of left-sided tumors, including rectum; however, the localization of the tumor in the right colon seems predominant compared to patients without MMR deficiency. Chapusot et al. reported a rate of 35% for right colon predominance in MMR-deficient tumors and emphasized that this tumor localization has a highly sensitive predictive value for poor tumor differentiation and MMR deficiency (29). The tumors with MSI-H phenotype tend to have high tumor invasion depth (pT) and low overall TNM stage. It has been widely accepted that this phenotype is associated with a low rate of lymph node metastasis (11). Similar to the literature, the present study found a significant relationship between MMR deficiency and pT and pN. In the present study, the rate of poorly differentiated tumors was significantly higher among MMR-deficient tumors than in MMR-stable tumors. This difference is particularly more remarkable in MLH1 and PMS2-deficient tumors. Reports indicate that patients with a loss of MMR protein expression often present with poorly differentiated and high-grade tumors. However, it's worth noting that this association has a low predictive value for predicting the loss of MMR protein expression (10,29). The fact that such tumors are still described by the Bethesda criteria mandates immunohistochemical screening of these tumors. The diagnosis of these tumors is not always straightforward, and the evaluation of these tumors in different categories in different studies complicates the interpretation of the results. For example, medullary tumors have an extremely low rate among all colorectal cancers. For this reason, medullary tumors are often included in the poorly differentiated tumor category in the studies (19). In our series, it has been problematic to differentiate undifferentiated or poorly differentiated tumors with medullary tumor-like areas from medullary tumors. It is a remarkable finding that one of the patients with medullary carcinoma in the present study had multiple tumors, a synchronous secondary tumor had mucinous adenocarcinoma histology, and two tumors had MLH1-PMS2 deficiency. Furthermore, the presence of a medullary tumor component was analyzed in a separate category isolated from patients with medullary carcinoma. Among patients with a medullary tumor

component, the rate of patients with loss of MMR protein expression was significantly higher than the rate of patients without MMR protein deficiency. In a study by Alexander et al., the presence of a medullary tumor component was reported in approximately 25% of patients with loss of MMR protein expression (22). It was emphasized that the presence of a medullary tumor component offers a specificity of 97% in predicting the loss of MMR protein expression (22). When MLH1-PMS2 and MSH2-MSH6-deficient tumors were evaluated separately, the rate of medullary tumor component was significantly higher in tumors without MMR protein deficiency. The mucinous tumor component is another phenotypic characteristic of MSI tumors. Many other studies have established that the loss of MMR protein expression is the most important predictive characteristic for the mucinous component (28,30). Also, in the present study, the rate of focal mucinous differentiation and mucinous adenocarcinoma was significantly higher among MMR-deficient tumors. In a study by Greenson et al. involving 528 patients, 29.1% of MMR-deficient tumors showed focal mucinous differentiation, and 28.6% showed more than 50% mucinous component (19). The same study also reported a sensitivity of 67.3% and a specificity of 81.9% for the presence of mucinous components in predicting MSI-H phenotype. The most remarkable histological characteristic of tumors with an MSI-H phenotype is the lymphocytic reaction to the tumor. TIL count is an important part of these lymphocytic reactions (18). There is no consensus in the literature regarding the method of determining the TIL count. The studies on the TIL count based on immunohistochemical studies using T-cell markers (CD3 or CD8) have found higher values than those using HE-stained preparations (19,22). Also, the cut-off value for TIL count that might be associated with MSI-H phenotype varies according to the method used. There are studies considering a cut-off value of 4-7 lymphocytes in immunohistochemical analysis (22,31), while the investigators relying on the H&E-stained slides have considered a cut-off value of 2-3 lymphocytes (19,32). Greenson et al. performed TIL counts using H&E staining and found that the most significant cut-off value is two lymphocytes associated with an MSI-H phenotype (19). They suggested that the presence of two or more lymphocytes in one high-power field offers a sensitivity of 90.4% and a specificity of 76.7% in predicting MSI-H phenotype (19). A cut-off value of two lymphocytes was used as recommended by Greenson et al (19). According to this analysis, TIL count was higher than 2 in 20% of patients with MMR protein deficiency and only in 8.3% of patients without MMR protein deficiency, and the difference was statistically significant. Crohn's-like lymphoid reaction is another lymphocytic reaction pattern (11,18). The rate of Crohn's-like lymphoid reaction was significantly higher in patients with MMR deficiency. In a study by Alexander et al. involving 204 patients, the rate of Crohn's-like lymphoid reaction was reported to be as high as 49% (22). They, however, emphasized that this feature is of low value in predicting MSI-H phenotype (29). It has been proposed that this reaction reflects the response of the host immune system to the tumor. It was also suggested that this reaction reduces the rate of metastasis and offers survival benefits to a certain extent (29). There were studies reporting that the presence of dedifferentiated cells (tumor budding) observed in some of the tumors with an infiltrative growth pattern might be an unfavorable prognostic parameter (20,33). However, tumor budding is more often observed in tumors without MMR protein deficiency (34). Also, in the present study, the rate of tumor budding was higher in patients without MMR protein deficiency. The absence of dirty necrosis is another histological parameter linked to MSI-H phenotype in recent studies (19). In a series of patients reported by Greenson et al., the absence of dirty necrosis offered a sensitivity of 82.7% and a specificity of 76.6% in predicting MSI-H phenotype (19). In the present study, the rate of dirty necrosis was 35.7% in MMR-deficient tumors, and the ratio difference between the two groups of tumors was statistically significant. It is known that MSI is a prognostic marker for survival in patients with colorectal cancer (10,11). Many studies have emphasized that the loss of MMR protein expression favorably affects disease-specific survival (10,28). In a study by Gafa et al., the impact of histological parameters (tumor diameter, localization, differentiation, mucin component, medullar characteristic, lymphocytic reaction, expansive growth pattern) and MSI status on disease-specific survival was investigated. It was found that MSI

status was the most crucial predictive factor, associated with increased disease-specific survival (35). Only overall survival data was available in the present study.

Limitations

Although the overall survival was longer in patients with MMR protein deficiency, no significant relationship was found between the loss of MMR protein expression and survival. The inconsistency with the literature data can be explained by the low rate of these patients in the entire study group and the fact that disease-specific survival data was not available in our study.

CONCLUSION

The characteristics associated with the loss of MMR protein expression include age under 50 years, advanced tumor stage, low number of metastatic lymph nodes, right colon localization, poor differentiation and high tumor grade, mucinous and medullary tumor component, increased TIL and Crohn's-like lymphoid reaction, and low rates of tumor budding and dirty necrosis. The clinicopathological findings of the present study are consistent with those reported in the literature. However, no significant relationship was found between MMR protein deficiency and survival, although survival was longer in patients with MMR-deficient tumors. Unlike many other studies, the present study evaluated medullary components in a separate category independently of poorly differentiated tumors. This distinction has shown that the presence of medullary tumor components is an important parameter in predicting the loss of MMR expression.

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References

1. Geiersbach KB, Samowitz WS. Microsatellite instability and colorectal cancer. *Arch Pathol Lab Med.* 2011;135:1269-77.
2. Maratt JK, Stoffel E. Identification of Lynch Syndrome. *Gastrointest Endosc Clin N Am.* 2022;32:45-58.
3. Jenkins MA, Hayashi S, O'Shea AM, Burgart LJ, Smyrk TC, Shimizu D, et al. Colon Cancer Family Registry. Pathology features in Bethesda guidelines predict colorectal cancer microsatellite instability: a population-based study. *Gastroenterology.* 2007;133:48-56.
4. Boland CR, Lynch HT. The history of Lynch syndrome. *Fam Cancer.* 2013;12:145-57.
5. Martínez-Roca A, Giner-Calabuig M, Murcia O, Castillejo A, Soto JL, García-Heredia A, et al. Lynch-like Syndrome: Potential Mechanisms and Management. *Cancers (Basel).* 2022;14:1115.
6. Chubak B, Heald B, Sharp RR. Informed consent to

- microsatellite instability and immunohistochemistry screening for Lynch syndrome. *Genet Med*. 2011;13:356-60.
7. Shia J. Immunohistochemistry versus microsatellite instability testing for screening colorectal cancer patients at risk for hereditary nonpolyposis colorectal cancer syndrome: Part I. The utility of immunohistochemistry. *J Mol Diagnostics*. 2008;10:293-300.
 8. Hampel H, Frankel WL, Martin E, Arnold M, Khanduja K, Kuebler P, et al. Feasibility of screening for Lynch syndrome among patients with colorectal cancer. *J Clin Oncol*. 2008;26:5783-8.
 9. Yoon YS, Yu CS, Kim TW, Kim JH, Jang SJ, Cho DH, et al. Mismatch repair status in sporadic colorectal cancer: immunohistochemistry and microsatellite instability analyses. *J Gastroenterol Hepatol*. 2011;26:1733-9.
 10. Rantanen P, Keränen A, Barot S, Ghazi S, Liljegren A, Nordenvall C, et al. The prognostic significance of microsatellite instability in colorectal cancer: a Swedish multi-center study. *Int J Colorectal Dis*. 2023;38:197.
 11. Mei WJ, Mi M, Qian J, Xiao N, Yuan Y, Ding PR. Clinicopathological characteristics of high microsatellite instability/mismatch repair-deficient colorectal cancer: A narrative review. *Front Immunol*. 2022;13:1019582.
 12. Sinicrope FA. DNA mismatch repair and adjuvant chemotherapy in sporadic colon cancer. *Nat Rev Clin Oncol*. 2010;7:174-7.
 13. Gelsomino F, Barbolini M, Spallanzani A, Pugliese G, Cascinu S. The evolving role of microsatellite instability in colorectal cancer: A review. *Cancer Treat Rev*. 2016;51:19-26.
 14. Kok M, Chalabi M, Haanen J. How I treat MSI cancers with advanced disease. *ESMO Open*. 2019;4:e000511.
 15. Brierley JD, Gospodarowicz MK, Wittekind C. *TNM Classification of Malignant Tumours*. 8th ed. Wiley-Blackwell; 2017.
 16. Carneiro F, Chan J, Cheung NYA. *WHO Classification of Tumours. Digestive System Tumours*. 5th ed. Lyon: IARC Press; 2019.
 17. Umar A, Boland CR, Terdiman JP, Syngal S, de la Chapelle A, Rüschoff J, et al. Revised Bethesda Guidelines for hereditary nonpolyposis colorectal cancer (Lynch syndrome) and microsatellite instability. *J Natl Cancer Inst*. 2004;96:261-8.
 18. Ogino S, Noshio K, Irahara N, Meyerhardt JA, Baba Y, Shima K, et al. Lymphocytic reaction to colorectal cancer is associated with longer survival, independent of lymph node count, microsatellite instability, and CpG island methylator phenotype. *Clin Cancer Res*. 2009;15:6412-20.
 19. Greenson JK, Bonner JD, Ben-Yzhak O, Cohen HI, Misevich I, Resnick MB, et al. Phenotype of microsatellite unstable colorectal carcinomas: Well-differentiated and focally mucinous tumors and the absence of dirty necrosis correlate with microsatellite instability. *Am J Surg Pathol*. 2003;27:563-70.
 20. Lugli A, Kirsch R, Ajioka Y, Bosman F, Cathomas G, Dawson H, et al. Recommendations for reporting tumor budding in colorectal cancer based on the International Tumor Budding Consensus Conference (ITBCC) 2016. *Mod Pathol*. 2017;30:1299-311.
 21. Joost P, Bendahl PO, Halvarsson B, Rambech E, Nilbert M. Efficient and reproducible identification of mismatch repair deficient colon cancer: Validation of the MMR index and comparison with other predictive models. *BMC Clin Pathol*. 2013;13:33.
 22. Alexander J, Watanabe T, Wu TT, Rashid A, Li S, Hamilton SR. Histopathological identification of colon cancer with microsatellite instability. *Am J Pathol*. 2001;158:527-35.
 23. Helderan NC, Andini KD, van Leerdam ME, van Hest LP, Hoekman DR, Ahadova A, et al. MLH1 Promotor Hypermethylation in Colorectal and Endometrial Carcinomas from Patients with Lynch Syndrome. *J Mol Diagn*. 2024;26:106-14.
 24. Shia J, Tang LH, Vakiani E, Guillem JG, Stadler ZK, Soslow RA, et al. Immunohistochemistry as first-line screening for detecting colorectal cancer patients at risk for hereditary nonpolyposis colorectal cancer syndrome: a 2-antibody panel may be as predictive as a 4-antibody panel. *Am J Surg Pathol*. 2009;33:1639-45.
 25. Perea J, Rodríguez Y, Rueda D, Marín JC, Díaz-Tasende J, Álvaro E, et al. Early-onset colorectal cancer is an easy and effective tool to identify retrospectively Lynch syndrome. *Ann Surg Oncol*. 2011;18:3285-91.
 26. Cescmecioglu Karavin E, Sağnak Yılmaz Z, Yazici H, Ersoz S, Mungan S. Comparison of Microsatellite Instability With Clinicopathologic Data in Patients With Colon Adenocarcinoma. *Cureus*. 2024;16:e57814.
 27. Bellizzi AM, Frankel WL. Colorectal Cancer Due to Deficiency in DNA Mismatch Repair Function. *Adv Anat Pathol*. 2009;16:405-17.
 28. Khoo JJ, Gunn A, Peh SC. Pattern of hMLH1, hMSH2 and hMSH6 expression and clinical characteristics in a sample of Malaysian colorectal carcinoma cases. *Malays J Pathol*. 2013;35:45-57.
 29. Chapusot C, Martin L, Mungra N, Rageot D, Bouvier AM, Bonithon Kopp C, et al. Sporadic colorectal cancers with defective mismatch repair display a number of specific morphological characteristics: relationship between the expression of hMLH1 and hMSH2 proteins and clinicopathological features of 273 adenocarcinomas. *Histopathology*. 2003;43:40-7.
 30. Kakar S, Aksoy S, Burgart LJ, Smyrk TC. Mucinous carcinoma of the colon: Correlation of loss of mismatch repair enzymes with clinicopathologic features and survival. *Mod Pathol*. 2004;17:696-700.

31. JASS JR. Pathology of Hereditary Nonpolyposis Colorectal Cancer. *Ann N Y Acad Sci.* 2006;910:62-74.
32. Ward R, Meagher A, Tomlinson I, O'Connor T, Norrie M, Wu R, et al. Microsatellite instability and the clinicopathological features of sporadic colorectal cancer. *Gut.* 2001;48:821-9.
33. Haddad TS, Lugli A, Aherne S, Barresi V, Terris B, Bokhorst JM, et al. Improving tumor budding reporting in colorectal cancer: a Delphi consensus study. *Virchows Arch.* 2021;479:459-69.
34. Kevans D, Wang LM, Sheahan K, Hyland J, O'Donoghue D, Mulcahy H, et al. Epithelial-mesenchymal transition (EMT) protein expression in a cohort of stage II colorectal cancer patients with characterized tumor budding and mismatch repair protein status. *Int J Surg Pathol.* 2011;19:751-60.
35. Gafà R, Maestri I, Matteuzzi M, Santini A, Ferretti S, Cavazzini L, et al. Sporadic colorectal adenocarcinomas with high-frequency microsatellite instability. *Cancer.* 2000;89:2025-37.